



Exploration de la biodiversité génétique des sols : programme Terragenome

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Microorganisms: « quantitative level »

- Microscopical Eukaryotes

- Prokaryotes

Whitman *et al.* 1998 PNAS, 95, 6578-6583. *Prokaryotes: The unseen majority.*

4-6 x 10³⁰ bacterial cells

1,2 x	10²⁹	Ocean
2,6 x	10²⁹	Soil
3,5 x	10³⁰	Deep soil
0,25 - 2,5 x	10³⁰	Ocean deep soil



Microorganisms: « qualitative level »



1 g of soil

Number of bacterial cells : 1 000 000 000

**Number of bacterial species:
10 000**

Torsvik 2002 (Science)

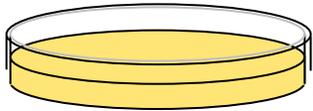
Roesch 2007 (ISME J.)

>10 000 000

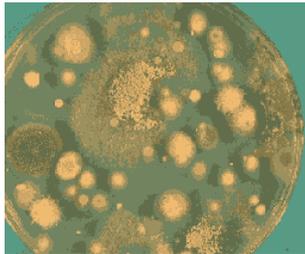
Gans 2005 (Science)



in vitro
Culture



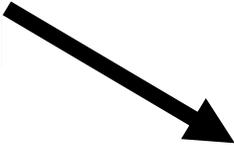
in vitro Isolation of
cultivable bacteria



Soil?

Percentage of cultivable bacteria?

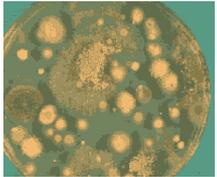
From 0,1 to 1%



(meta-) genomics approach

«Environmental DNA»

in vitro
Culture



***in vitro* culture approach**



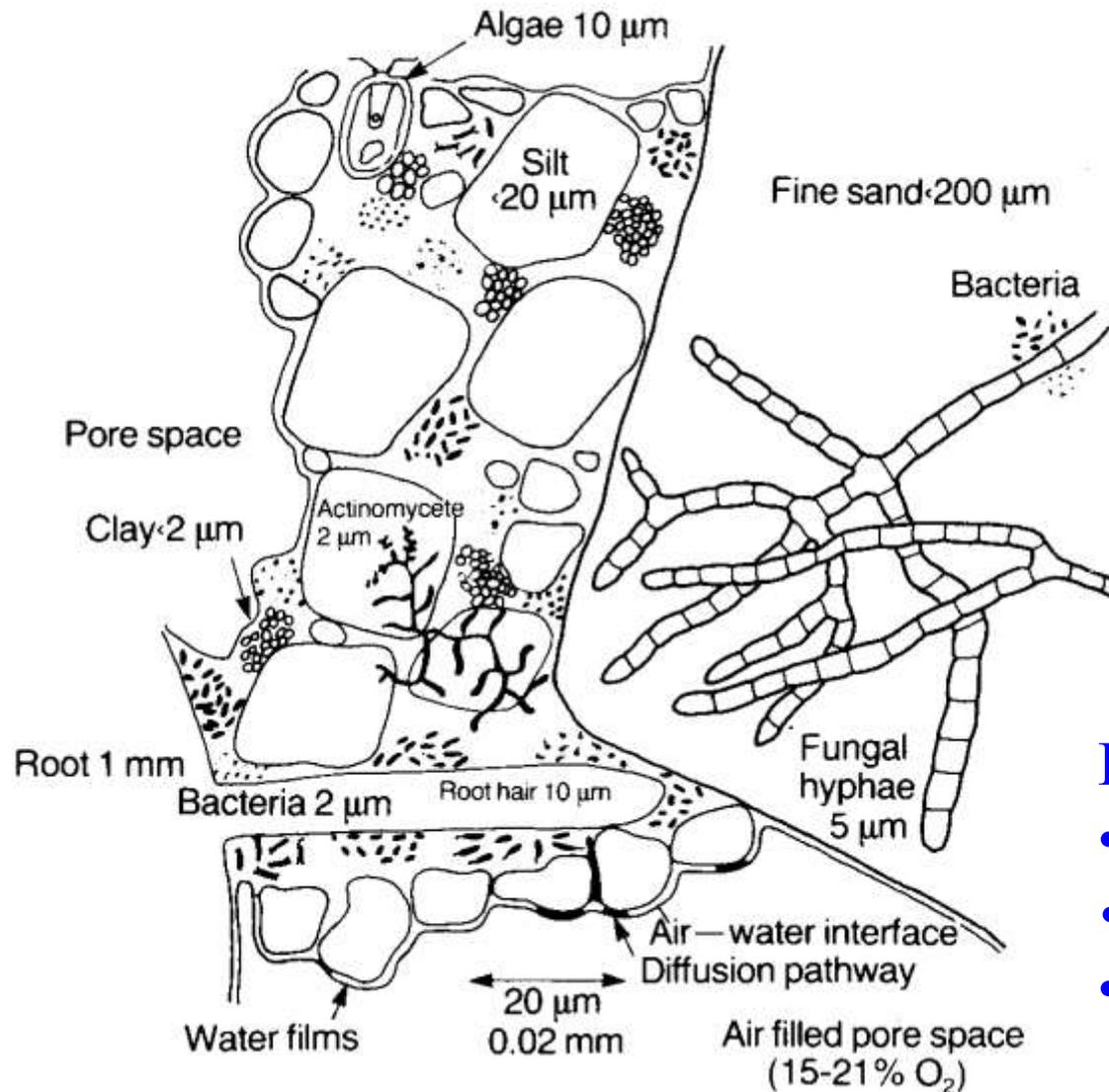
Cultivable bacteria? From 0,1 to 1%



Soil, a Composite of Communities

Its very diverse
Complex gradients
Unmixed

- microaggregates
- rhizosphere
- fungalsphere
- fauna
- pore surfaces
- OM coatings

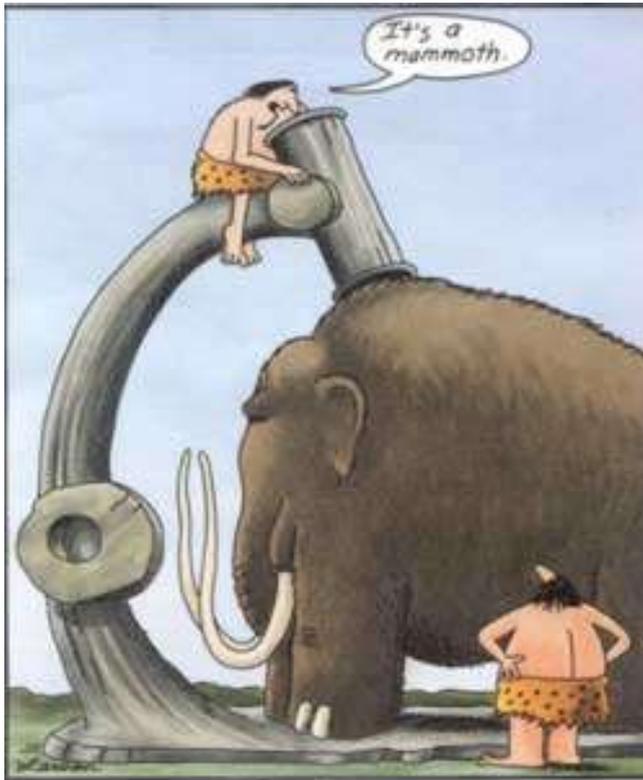


Implications:

- *Spatial isolation*
- *Minimizes competition*
- *How to sample since multiple communities?*

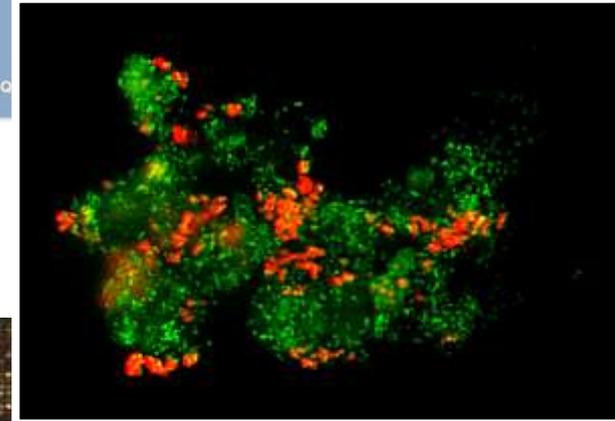
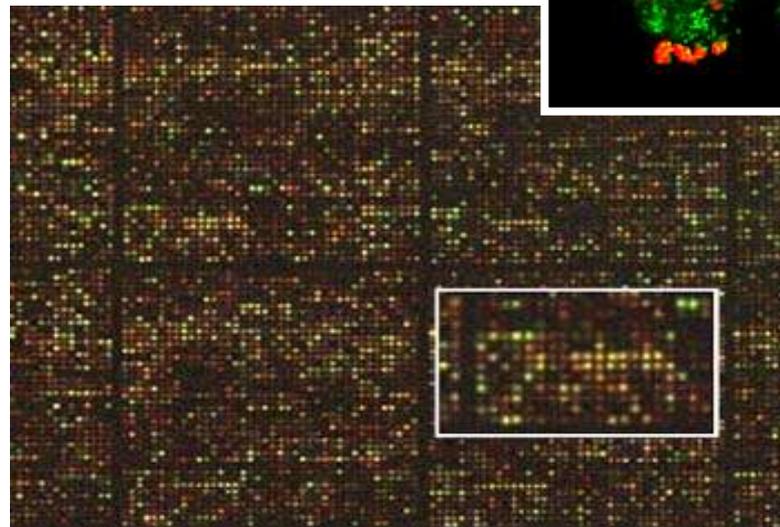
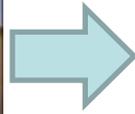
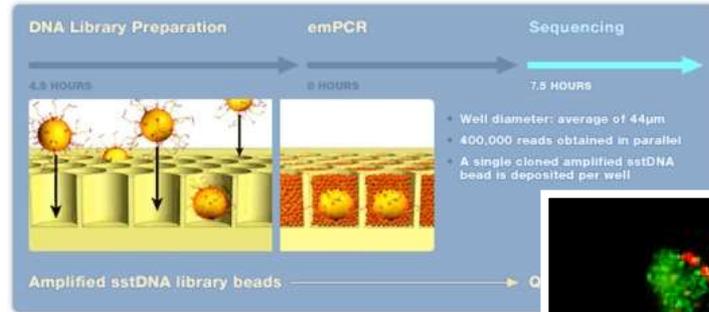


How 30 years has changed our technology to perceive life on Earth



Early microscopes

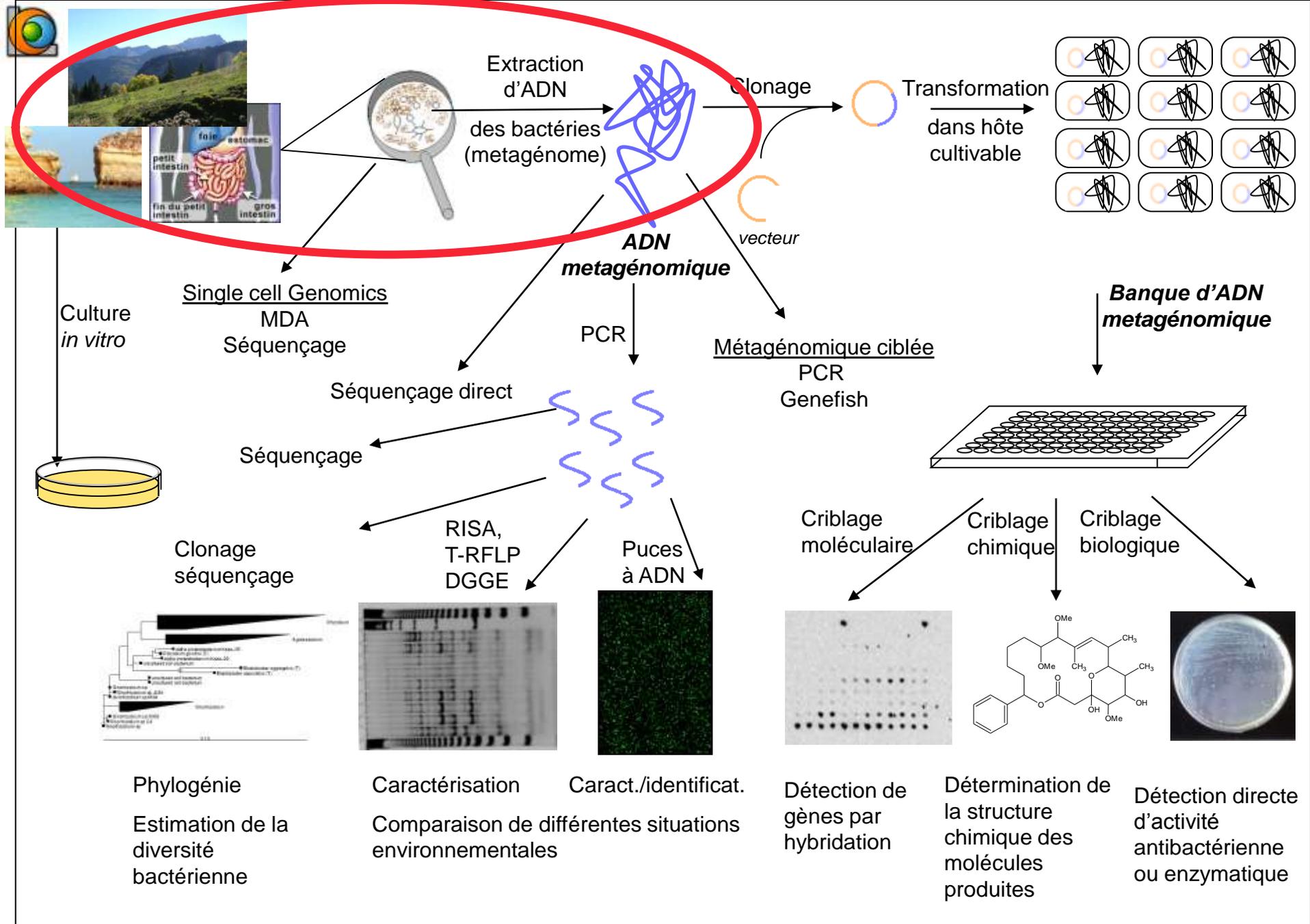
FIGURE 9





De nombreuses raisons de s'intéresser aux communautés bactériennes

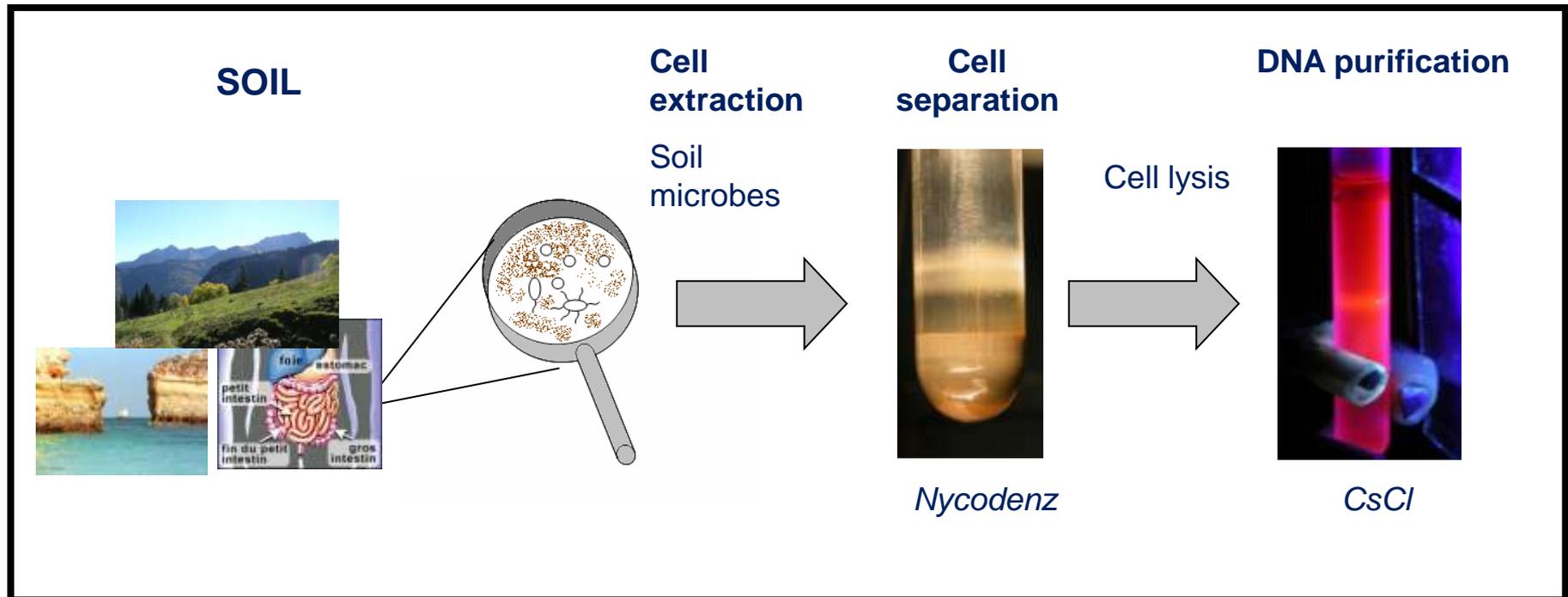
- impact sur les équilibres biogéochimiques
 - quels sont les acteurs ?
 - nouvelles étapes des cycles biologiques des éléments
- impact sur la santé (flores microbiennes humaines)
- modèles d'écosystèmes (structure des communautés bactériennes)
- utilisation de la biodiversité à des fins d'applications
 - substances thérapeutiques
 - substances d'intérêt industriel
 - enzymes utiles pour la chimie de synthèse
 - bioremédiation
- nouveaux éclairages sur l'évolution





Metagenome DNA extraction

- *In situ* lysis and total DNA extraction
- Cell extraction and lysis





Optimization of bacterial DNA recovery

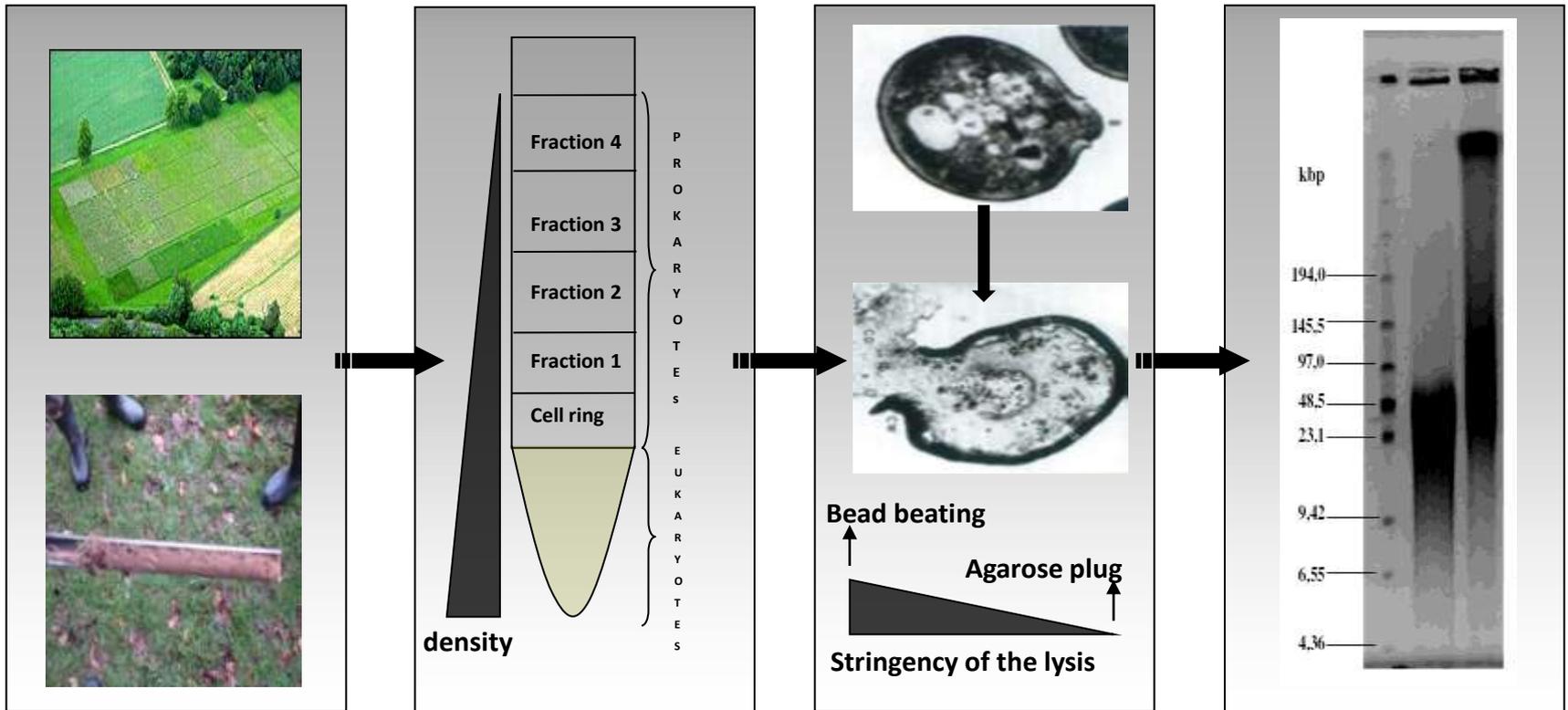
Sampling strategies

- Time of the year
- Depth

Improvement of cell recovery (Nycodenz)

Improvement of DNA recovery (sensitivity to lysis treatments)

Improvement of DNA recovery (DNA degradation)





Phylochip probes

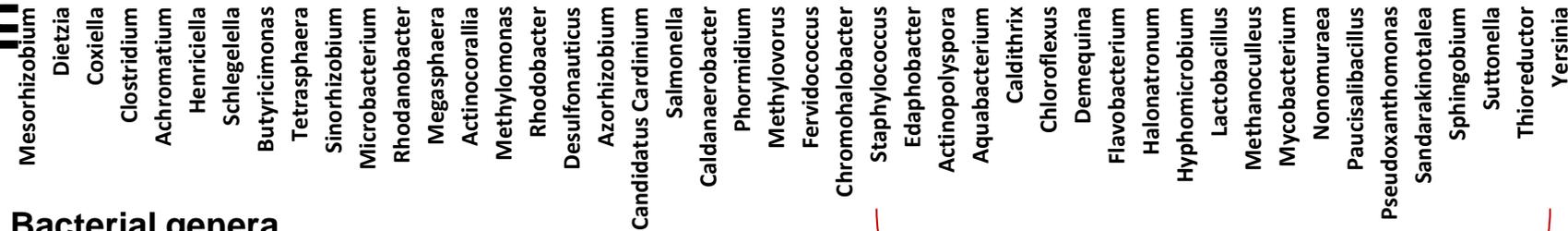
intensity

25

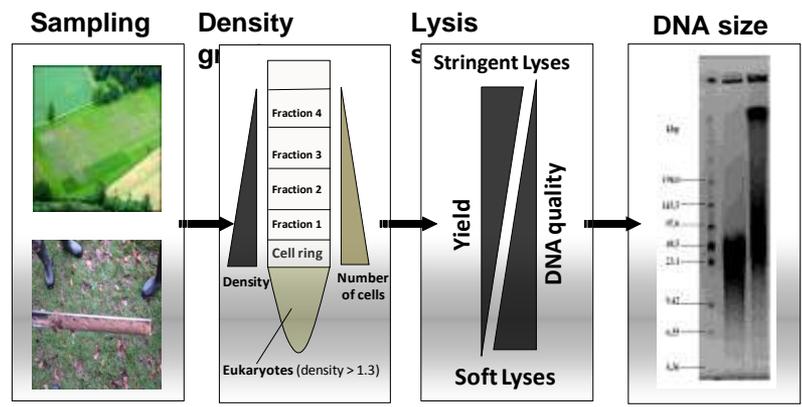
20

15

10



Bacterial genera

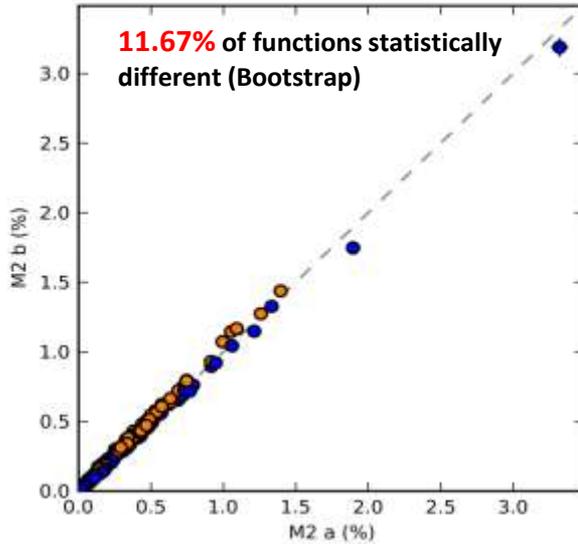


Undetected with one DNA extraction method

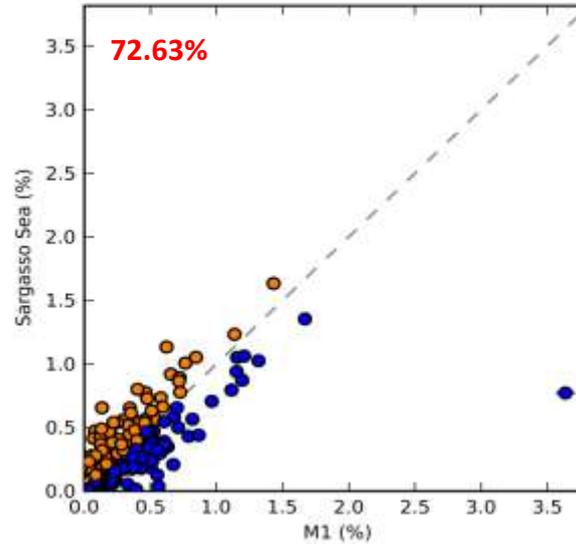


Functional comparison using MG RAST annotation and STAMP statistical analyses

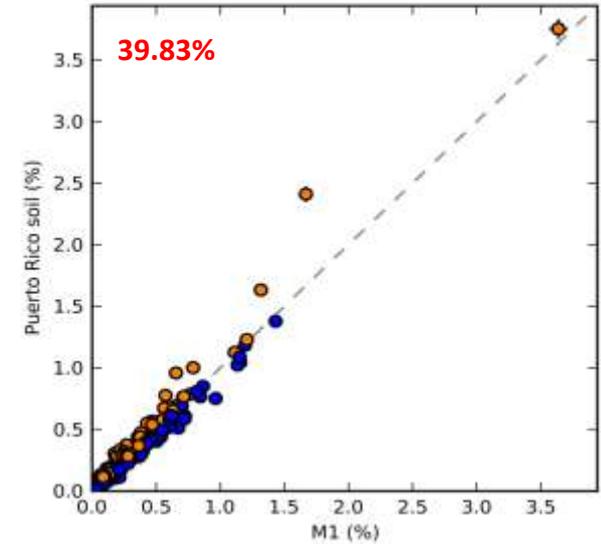
1. technological reproducibility



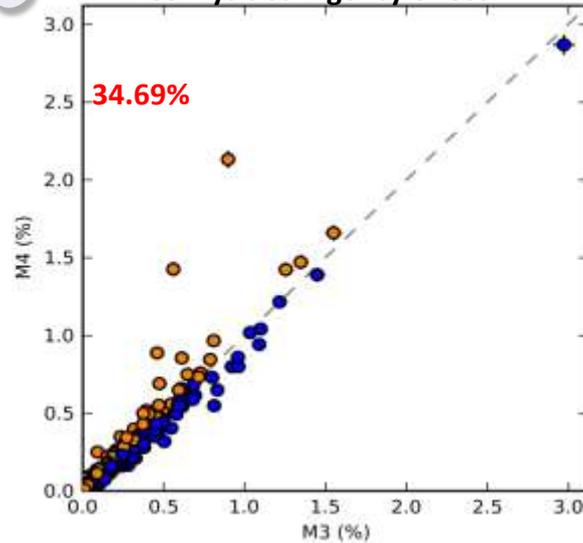
2. comparison with an ocean



3. comparison with another soil

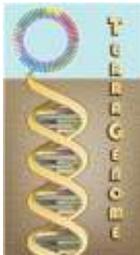


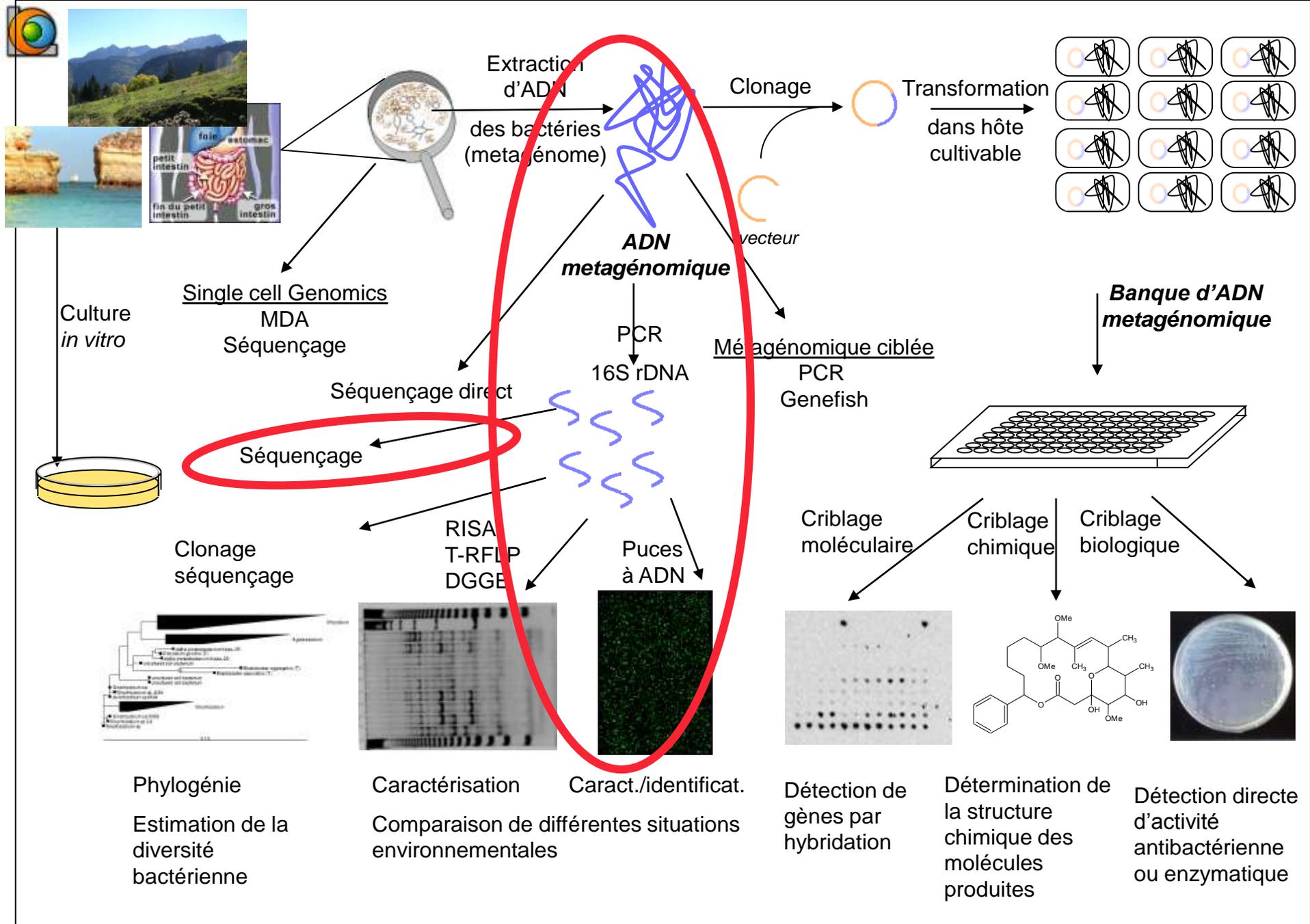
4. Cell lysis stringency effect



DNA Extraction bias

Functional level







Progression of the Sequencing Era

First generation (1995- 2007)

<u>Instrument</u>	<u>Length</u>	<u>Nos</u>	<u>Output</u>	<u>Time/run</u>
Sanger	700bp	96	60Kb	

Second generation (2006 - now)

Roche/titanium	400bp	1 mil	400Mb	1-2da
Illumina GAII	75bp x 2	30mil	2.2Gb	8da
Illumina HiSeq	100bp	1-2bil	200 Gb	8da
ABI SOLiD	75bp	3-4 bil	300Gb	14 da

Third generation (single molecule read) (late 2010)

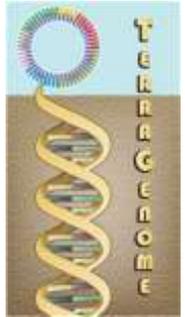
PacBio	>1000bp	0.1-0.3bil	1Gb	15 min
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Metasoil Project



The initial support for Terragenome (complete sequencing of a reference soil metagenome) :



Objective:

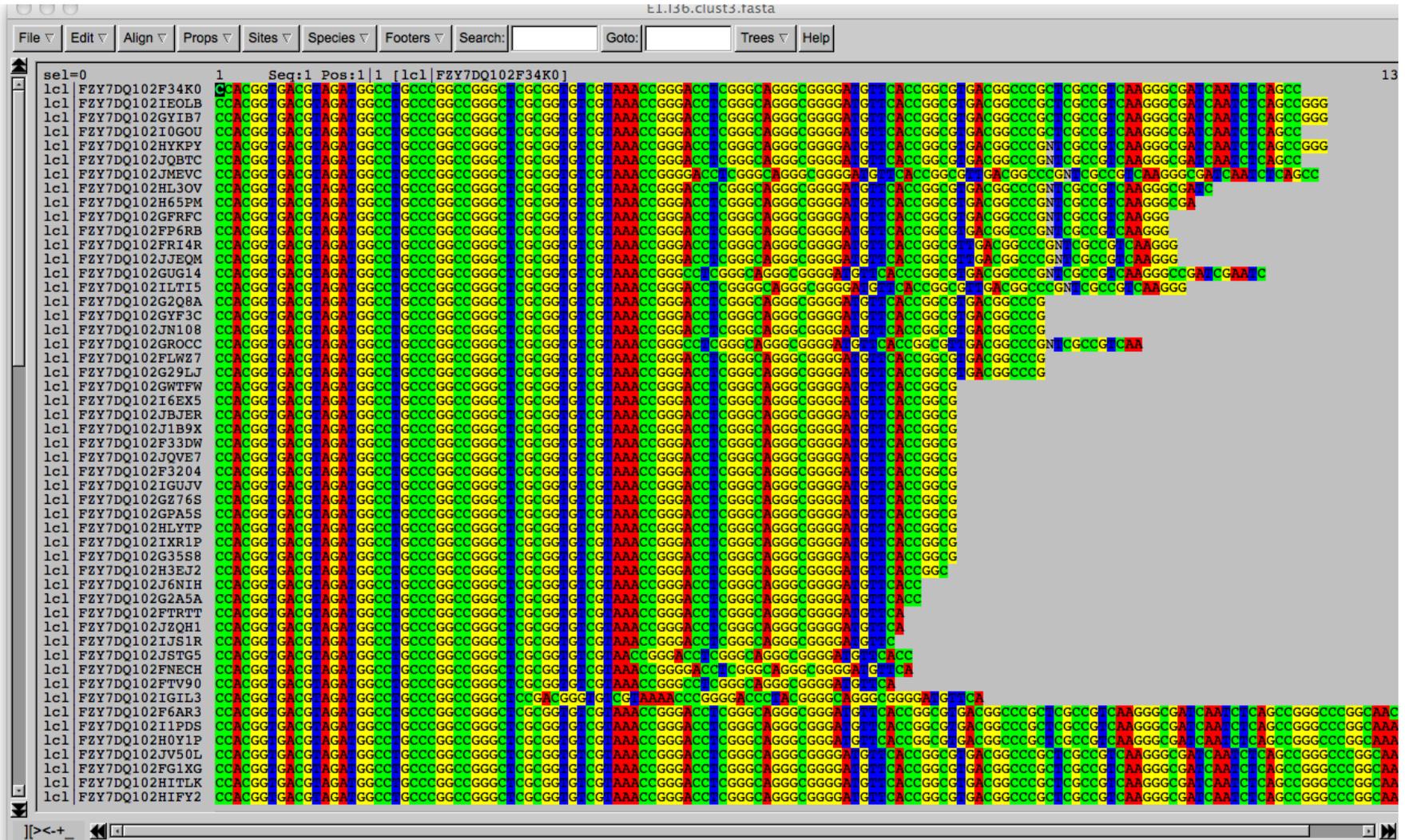
- Optimization of bacterial DNA recovery.
- Metagenomic DNA library construction
- Pyrosequencing of directly extracted DNA



Park Grass, Rothamsted: an internationally recognized agroecology field experiment for 150 years



Too many copies



Use cd-hit-454

Niu et al. Artificial and natural duplicates in pyrosequencing reads of metagenomic data. BMC Bioinformatics (2010) vol. 11 pp. 187

A middle-aged man with short dark hair and a goatee is smiling and holding a large, rectangular sign made of flattened cardboard. The sign is held in front of his chest and has the words "TOO MUCH DATA-NEED HELP!" written on it in red marker. He is wearing a dark brown jacket over a grey turtleneck sweater and a plaid scarf. The background shows a paved street with a crosswalk, a person walking in the distance, and bare trees under a clear sky. The sign is the central focus of the image, with the text written in a simple, hand-drawn style. The man's expression is positive, suggesting a lighthearted or humorous take on the message.

TOO MUCH
DATA-NEED
HELP!



We are now in a position to generate huge quantities of sequence (and other) information ... but are we traveling efficiently?

Bulk data
generation
capacity



Screening &
bioinformatic
strategies



We are now in a position to generate huge quantities of sequence (and other) information ... but are we traveling efficiently?

Technology

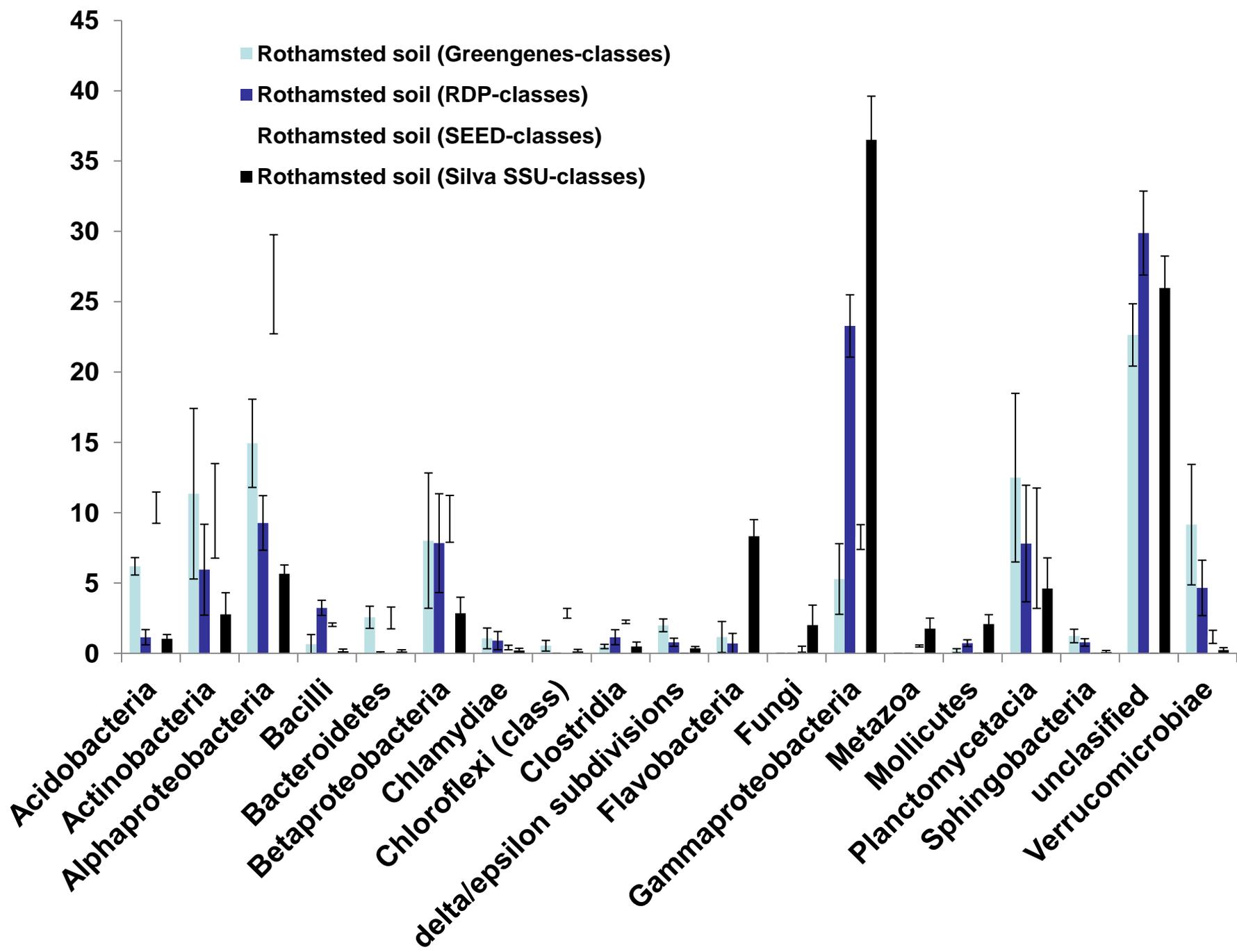


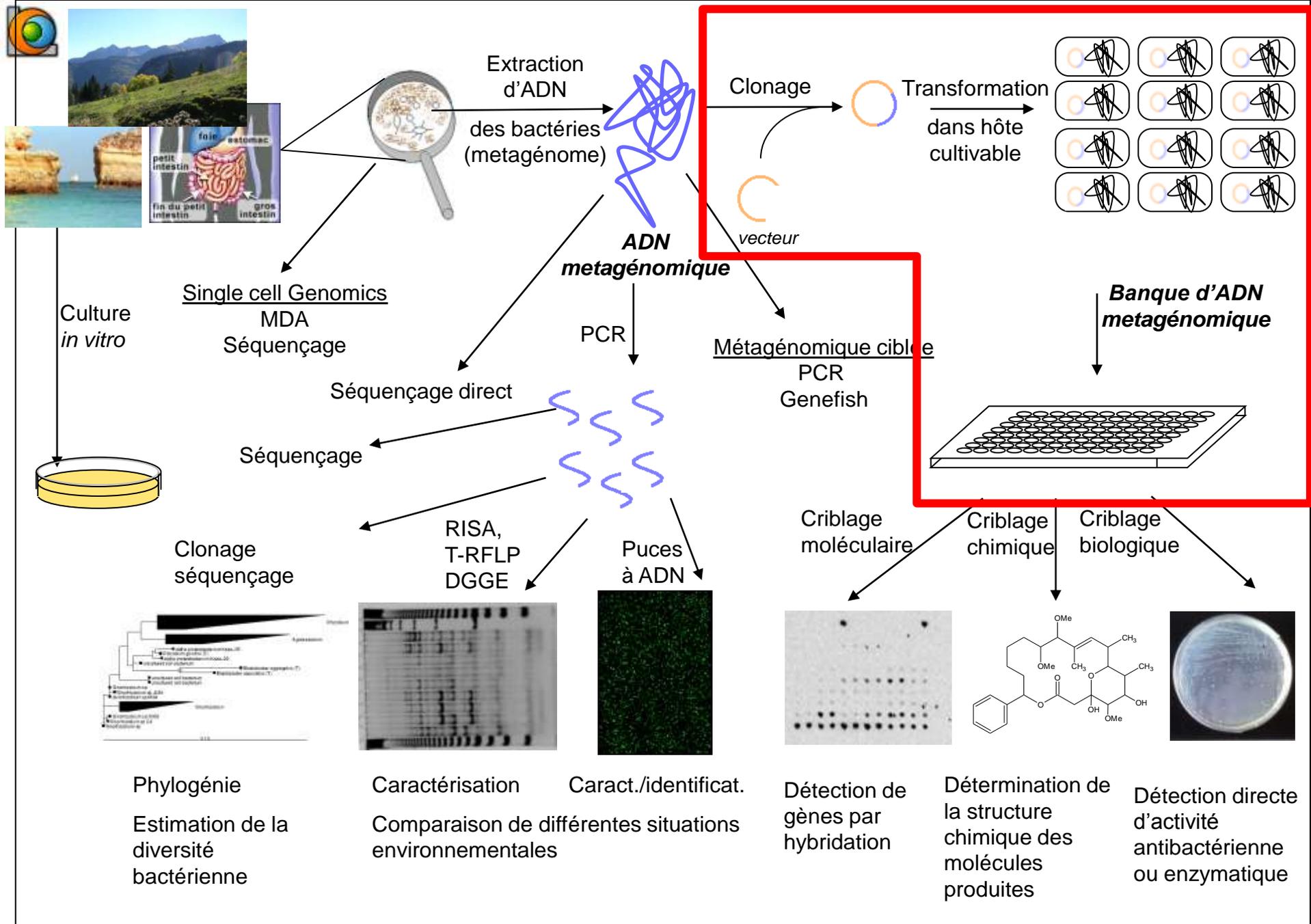
Science

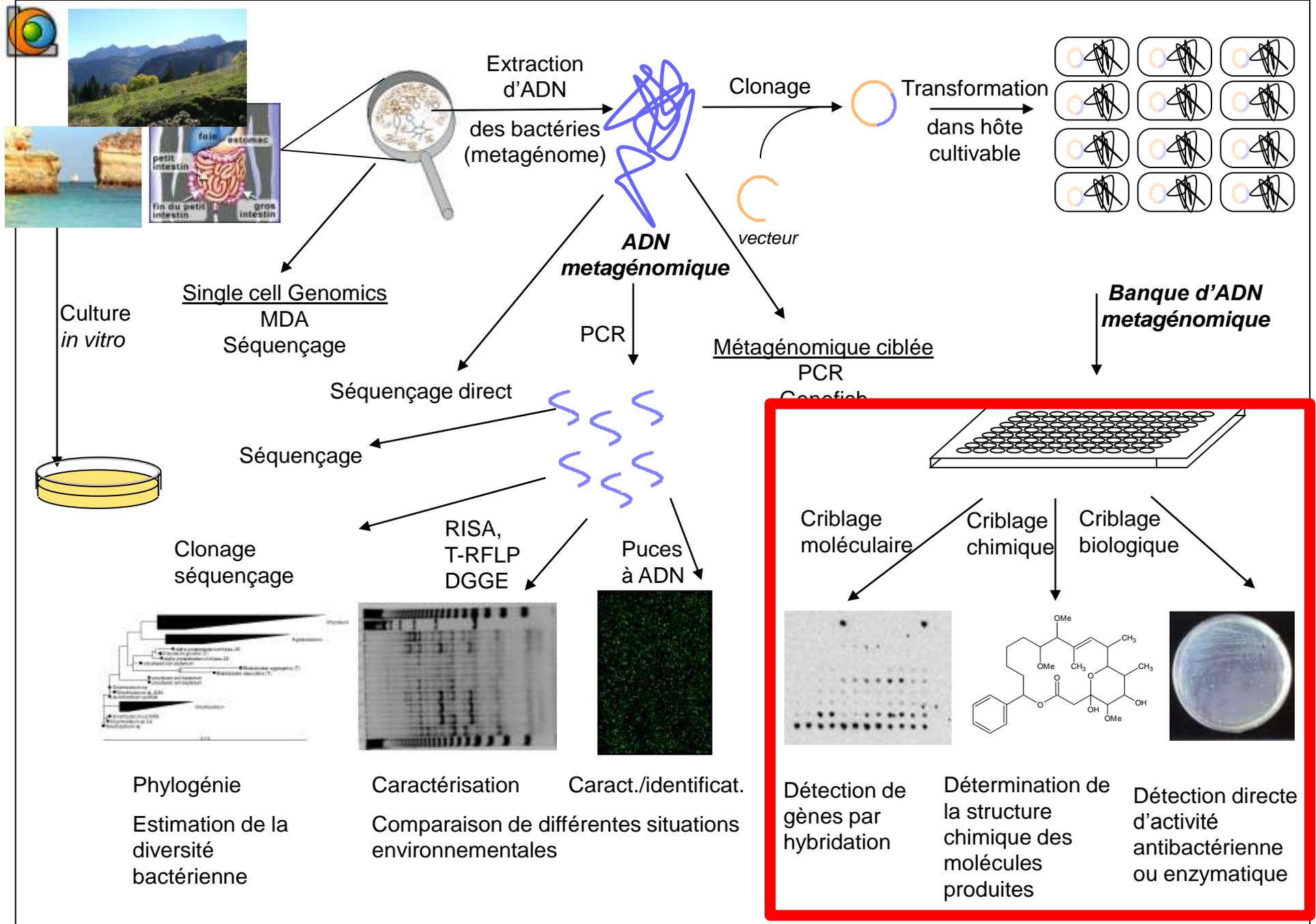
Courtesy of Jim Prosser and George Kowalchuk



HAVE. SOFTWARE
NEED DATA

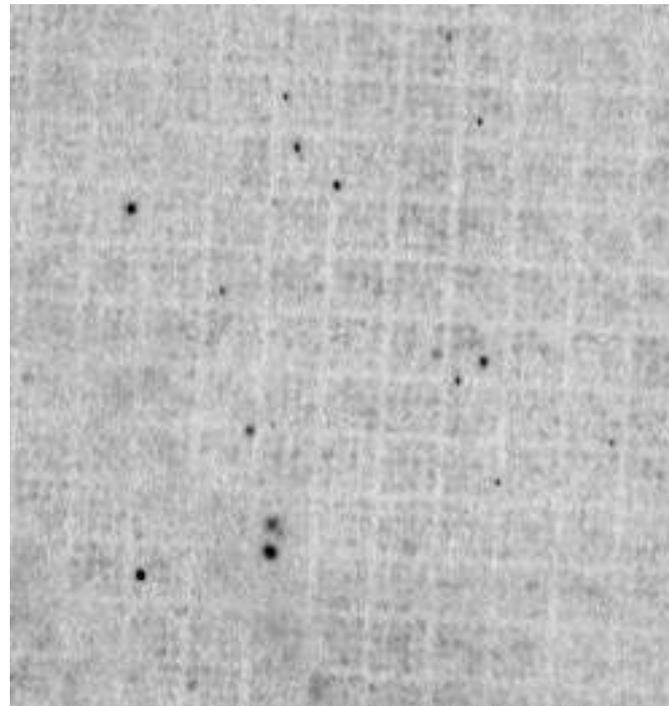








Molecular screening (60 000 clones)





Molecular screening

- 60 000 clones 139 « KS » +++
- 40 « KS »: sequencing No redundancy
Novelty
(similarity < 67%)



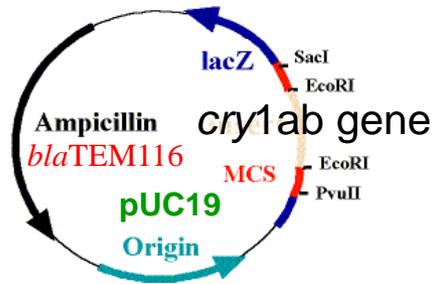
La métagénomique appliquée à la problématique des plantes OGM !



Experimental station: Bazièges, France

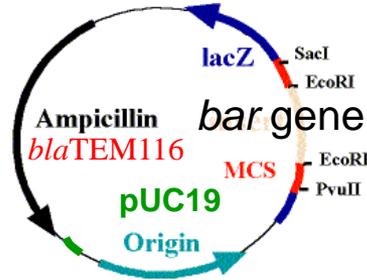


pCIB4431



Zea mays L.

pCIB3064



Event Bt176

Field with transgenic Bt 176 corn
(*bla*TEM-116) 10-years culture



Ampicillin-resistant bacteria isolation and bacterial metagenomic DNA extraction

Beta-lactam resistance tests

Controls with a traditional corn field and a prairie in the same location

Diversity of *bla*TEM genes
(PCR + cloning)
Comparisons of bacterial communities
(microarray data)



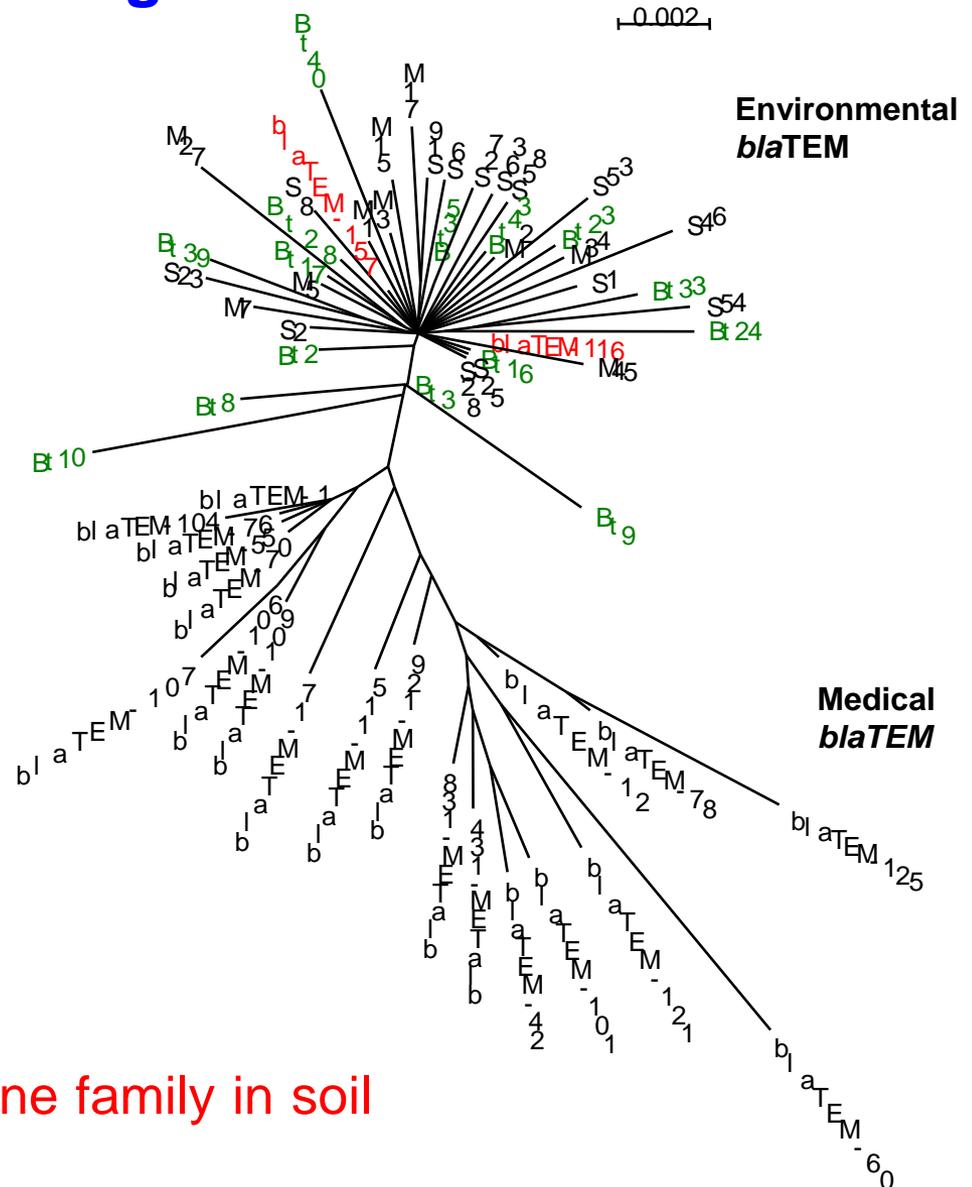
Diversity of *bla*TEM genes in soils

Metagenomic approach
=
DNA extracted from soil

10 DNA inserts that originated from transgenic and prairie soil DNA exhibited the *bla*TEM116 gene sequence, confirming its natural occurrence in environmental bacteria.

153 different *bla*TEM sequences were found in the PCR-amplified bacterial DNA.

High polymorphism level of this gene family in soil bacterial populations





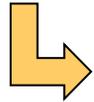
Transfert de gènes des plantes (OGM) aux bactéries?

- Est-ce que l'acquisition d'ADN étranger par des bactéries est possible?
- Si oui, est-ce qu'il y a une régulation de l'acquisition de l'ADN?
Par quels mécanismes?
- Est-ce que l'ADN de la plante peut être transféré aux bactéries?
- Est-ce qu'il y a une spécificité pour l'ADN des plantes OGM ?
- Est-ce que l'on peut détecter un transfert de gènes des plantes vers les bactéries?
Dans quels environnements? A quelle fréquence?
- **Est-ce qu'il y a un impact sur la communauté des bactéries?**



Conclusions

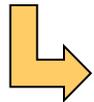
√ Indigenous bacteria possess a wide diversity of beta-lactam resistance genes



Involvement in evolutionary processes

√ Soil is a reservoir of genetic determinants providing bacteria with the ability to adapt rapidly to present and future antibiotics!

√ Acquisition of *bla* gene from transgenic plants would not modify existing gene diversity in soil nor confer specific advantage



The risk that antibiotic resistances in GMP can pose to clinical strains could be considered as almost null



Aurélie Faugier, Sébastien Cécillon,
Davide Francioli, Tom Delmont
Emmanuel Prestat, Jean-Michel
Monier, Timothy M Vogel,

Environmental
Microbial
Genomics



www.GenomEnviron.org